

# Thinking too positive? Revisiting current methods of population genetic selection inference

Claudia Bank<sup>1,2</sup>, Gregory B. Ewing<sup>1,2</sup>, Anna Ferrer-Admettla<sup>1,2,3</sup>, Matthieu Foll<sup>1,2</sup>, and Jeffrey D. Jensen<sup>1,2</sup>

In the age of next-generation sequencing, the availability of increasing amounts and improved quality of data at decreasing cost ought to allow for a better understanding of how natural selection is shaping the genome than ever before. However, alternative forces, such as demography and background selection (BGS), obscure the footprints of positive selection that we would like to identify. In this review, we illustrate recent developments in this area, and outline a roadmap for improved selection inference. We argue (i) that the development and obligatory use of advanced simulation tools is necessary for improved identification of selected loci, (ii) that genomic information from multiple time points will enhance the power of inference, and (iii) that results from experimental evolution should be utilized to better inform population genomic studies.

# Identification of beneficial mutations in the genome: an ongoing quest

The identification of genetic variants that confer an advantage to an organism, and that have spread by forces other than chance, remains an important question in evolutionary biology. Success in this regard will have broad implications, not only for informing our view of the process of evolution itself, but also for evolutionary applications ranging from clinical to ecological. Despite the tremendous quantity of polymorphism data now at our fingertips, which, in principle, ought to allow for a better characterization of such adaptive genetic variants, it remains a challenge to unambiguously identify alleles under selection. This is primarily owing to the difficulty in disentangling the effects of positive selection from those of other factors that shape the composition of genomes, including both demography as well as other selective processes.

Approaches to identify positively selected variants from genomic data can be broadly divided into two categories: those that make use of within-population polymorphism

Corresponding author: Bank, C. (claudia.bank@epfl.ch).

Keywords: natural selection; background selection; population genetic inference; evolution; computational biology.

0168-9525/

© 2014 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.tig.2014.09.010

data, and those that make use of between-population/species data. While each approach has its respective merits, in this review, we focus on recent developments in population genetic inference from polymorphism data in both natural and experimental settings (see [1,2] for more general reviews, and [3,4] for recent and specific literature on divergence-based selection inference). For population genetic inference from single time point polymorphism data (as is most commonly the case), this includes not only sophisticated statistical methods, but also simulation programs that enable us to model expected genomic signatures under a wide variety of possible scenarios.

### Glossary

**Background selection:** reduction of genetic diversity due to selection against deleterious mutations at linked sites.

**Coalescent simulator:** simulation tool that reconstructs the genealogical history of a sample backwards in time. This greatly reduces computational effort, but only models in which mutations are independent of the sample's genealogy can be implemented.

Cost of adaptation: the deleterious effect that a beneficial mutation can have in a different environment. Prominent examples are antibiotic resistance mutations, which have often been observed to cause reduced growth rates (as compared with the wild type) in the absence of antibiotics.

**Demographic history:** the population history of a sample of individuals, which can include population size changes, differing sex ratios, migration rates, splitting and reconnection of the population, as well as variation over time in these parameters.

**Distribution of fitness effects (DFE):** the statistical distribution of selection coefficients of all possible new mutations, as compared with a reference genotype.

**Epistasis:** the interaction of mutational effects, resulting in a dependence of the effect of a mutation on the background it appears on.

Forward simulator: simulation tool that models the evolution of populations forward in time. This allows for implementation of complex models, but also usually results in much longer computation times because all individuals/haplotypes must be tracked.

**Nonequilibrium model:** any model that incorporates violations of the assumptions of the standard neutral model (see below).

**Selection coefficient:** a measure of the strength of selection on a selected genotype. Usually, the selection coefficient is measured as the relative difference between the reproductive success of the selected and the ancestral genotypes.

**Selective sweep:** the process of a beneficial mutation (and its closely linked chromosomal vicinity) being driven ('swept') to high frequency or fixation by natural selection. Selective sweeps result in a genomic signature including a local reduction in genetic variation, and skews in the SFS.

**Standard neutral model:** under this model [67], the population resides in an equilibrium of allele frequencies determined by the (constant) mutation rate and population size. The model assumptions include random mating, binomial sampling of offspring, and no selection.



<sup>&</sup>lt;sup>1</sup> School of Life Sciences, Ecole Polytechnique Fédérale de Lausanne (EPFL), 1015 Lausanne, Switzerland

<sup>&</sup>lt;sup>2</sup> Swiss Institute of Bioinformatics (SIB), 1015 Lausanne, Switzerland

<sup>&</sup>lt;sup>3</sup> Department of Biology and Biochemistry, University of Fribourg, 1700 Fribourg, Switzerland

Alternatively, data from multiple time points – such as those recently afforded by ancient genomic data as well as many clinical and experimental datasets – can be used to greatly improve inference by catching a selective sweep 'in the act'. Finally, recent results from the experimental evolution literature have begun to better illuminate the expected distribution of fitness effects (DFE), the associated costs of adaptation, and the extent of epistasis (see Glossary). In this opinion piece, we present an overview of recent developments for selection inference in the abovementioned areas, and offer a roadmap for future method development.

# Selection inference from a single time point

One of the earliest efforts to quantify selection in a natural population was based on multiple time point phenotypic data [5,6]. At the onset of genomics, however, new sequencing methods were both tedious and expensive, limiting data collection to single time points. For this reason, one generally observes only the footprints of the selection process, making it more difficult to distinguish regions shaped by neutral processes from those shaped by selective processes. Over the past number of decades, population genetic theory has predicted the effects of different selection models on molecular variation. These predictions have given rise to test statistics designed to detect selection using polymorphism data, based on patterns of population differentiation (e.g., [7–10]), the shape of the site-frequency spectrum (SFS) (e.g., [11–13]), and haplotype-linkage disequilibrium (LD) structure (e.g., [14-17]).

Despite efforts to create statistics robust to demography, all currently available methods to detect selection are prone to misinference under nonequilibrium models

[18,19]. Therefore, in parallel to the production of statistics for inferring selection, a separate class of methods has been developed to estimate the demographic history of populations utilizing the same patterns of variation [20,21]. This lends itself to a two-step approach when analyzing population genetic data: first, demography is inferred using a putatively neutral class of sites, and that model is then used to test for selection among a putatively selected class of sites [13,16]. However, the assumptions enabling this inference are highly problematic because they rely on the correct identification of a class of sites that is both neutral and untouched by linked selection. BGS, for example, may indeed influence a large fraction of the genome in many species [22]. Thus, the misidentification of such a class of sites in the genome may not only result in the misinference of the underlying demographic history, but also in misinference of selection owing to the incorrect estimation of the demographic null model leading to both false positives and false negatives (Figure 1).

In order to circumvent this problem, it is, therefore, necessary to develop methods that can jointly infer the demographic and selective history of the population simultaneously, recognizing that both processes are likely to shape the majority of the genome in concert [23]. The development of simulation software that can model both positive and negative selection in nonequilibrium populations (see below) is a step towards this goal, as it allows for the generation of expected patterns of variation under such scenarios. Recently, the introduction of likelihood-free inference frameworks such as Approximate Bayesian Computation (ABC) [24] has made it computationally feasible to combine demographic and selective inference. Recent

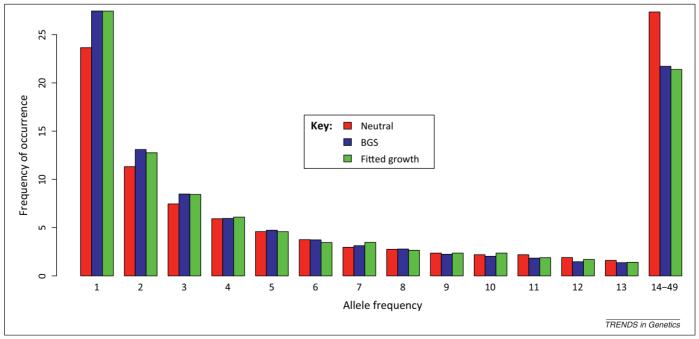


Figure 1. An example of the ability of background selection to mimic neutral nonequilibrium (here, exponential growth) site-frequency spectrum (SFS) based patterns. Assuming an absence of background selection (BGS) effects when they are present (as is common in demographic inference), may result in substantial misinference of the underlying demographic model – here inferring population growth when the population is, in fact, at equilibrium. Simulations were performed using SFSCode [40], with 50 samples from a single time point, conditioned on 100 single nucleotide polymorphisms per locus. The BGS coefficient is  $\alpha = 2Ns = -4$  and the probability of a deleterious mutation is 0.1, with recombination rate  $\rho = 2Nr = 50$  between chromosome ends. The single nucleotide polymorphisms (SFS) shows the frequency of occurrence (y-axis) of a number of derived alleles (x-axis) in the simulated sample. Site classes 14–49 were binned for illustrative purposes.

advances in this field allow for management of the large number of summary statistics needed to achieve this goal [25,26], and the pieces appear largely in place for significant progress to be made on this front.

# Selection inference from multiple time points

As noted above, although temporal data was considered in the infancy of population genetics, it was not until the late 1980s that multitime point genetic datasets became available, spurring the development of related statistics. These approaches focused first on the estimation of changes in population size [27–30]; later, new methods were developed specifically for time serial data to co-estimate parameters such as the intensity of selection and the effective population size [31].

Multitime point methods have a major advantage over single time point based approaches in that knowledge of the trajectory of an allele provides valuable information about the underlying selection coefficient. Owing to advances in sequencing technologies, there is now an increased availability of multitime point data of not only an experimental, but also a nonexperimental nature (ranging from ancient samples to those from longitudinal medical studies or field work). This temporal dimension spurred the development of multiple time point based methods over the last few years – all seeking to estimate some combination of selection coefficient and (i) effective population size [32,33], (ii) migration [34], (iii) the age of the selected mutation [31], or (iv) the recombination rate [35]. These methods differ both in the underlying models and in their respective performance, and hence in the conditions to which they are best suited (Table 1; Table S1 in the supplementary material online). In Malaspinas et al. [31] and Mathieson and McVean [34], the trajectory of the selected allele is modeled as a hidden Markovian process, and the observations are represented as binomial observations from the population. Subsequently, Malaspinas et al. [31] use an approximate transition density to compute the likelihood, whereas Mathieson and McVean [34] use an expectation-maximization algorithm to maximize the likelihood, which speeds up the estimation procedure. By contrast, Foll *et al.* [32,33] do not use a Hidden Markov Model (HMM), but an ABC approach to obtain the posterior distribution of the selection coefficient after simulating allele frequency trajectories using the Wright–Fisher model under a range of selection coefficients. This approach results in lower accuracy for very small selection coefficients, but also represents the fastest and least biased method to date.

Although current multitime point methods are better at inferring the parameters of positive selection than single time point methods, a number of challenges remain. Firstly, maximum likelihood estimators (MLE) may have higher accuracy for small selection coefficients than an ABC-based approach, but currently available implementations are extremely computationally intensive and thus cannot be readily used as scans of selection (i.e., a candidate site must be known a priori – a generally unlikely scenario), and the number of time points necessary to achieve sufficient power still remains largely unexplored. In addition, owing to the recent development of these methods, a relatively small number of selection models have been considered – primarily those of consistent and directional positive or negative selection acting on the candidate site. As such, they suffer from the same limitation described above for single time point inference in that they are unable to jointly estimate selection and the demographic history of the population. Secondly, such methods have, thus far, been unable to effectively tease apart the effects of direct versus linked selection. By contrast, given the computational efficiency of recently proposed ABC-based approaches [32,33], it has indeed become possible to effectively utilize such approaches in order to identify candidate sites from whole genome polymorphism data, thereby avoiding the first limitation. In addition, because of its efficiency and ability to handle multiple summary statistics, ABC appears as the most promising framework in which to consider more complex selection models under nonequilibrium demographic scenarios.

Table 1. An overview of currently available multiple time point selection estimators and their strengths and weaknesses

Author	Refs	Type	Advantages	Disadvantages	
Bollback et al. (2008)	[68]	MLE		Severely biased.	
Malaspinas <i>et al.</i> (2012)	[31]	MLE	Can estimate allele age. Very accurate for small s values.	Slow, in particular for large $N_e$ s values. Biased for large s values.  Assumes that the allele is segregating at the last sampling point, leading to some bias in other scenarios. $N_e$ poorly estimated independently at each locu	
Mathieson and McVean (2013)	[34]	MLE	Fast.  Jointly estimates migration rate and spatially varying selection coefficients in different populations.	$N_e$ assumed known. No model of ascertainment implemented, leading to some bias under realistic scenarios. $N_e$ poorly estimated independently at each locus.	
Foll <i>et al.</i> (2014)	[33]	ABC	<ul> <li>N<sub>e</sub> estimated using all loci together.</li> <li>Very fast.</li> <li>Unbiased for whole range of s.</li> <li>Can estimate dominance level in some cases.</li> <li>Wide range of ascertainment scenarios implemented.</li> </ul>	$N_e$ estimate assumes most loci are neutral. Not as accurate as MLE methods for small $s$ .	

<sup>&</sup>lt;sup>a</sup>Abbreviations: N<sub>e</sub>, effective population size; s, selection coefficient.

#### Simulating selection

Simulation programs have long been an important tool in population genetics, both for validating theoretical work and for characterizing real data. In the area of selection inference, simulations are used to investigate appropriate critical values of test statistics under nontrivial models and for detecting deviations from the standard neutral model [11,36,37]. Recently, the importance of efficient simulation under complex models has become relevant both for understanding expected patterns of variation and for direct inference using likelihood free ABC [24].

Currently, there are a number of simulation programs available that can simulate a wide range of scenarios and are applicable to different types of models and data (see Table 2). Broadly speaking, simulation programs can be split into two different types. Forward simulators can include a wide range of demographic and selective models [38-43]. However, because every gamete needs to be tracked and every generation simulated, they tend to be computationally costly and thus slow. By contrast, coalescent simulators are very fast as only the sample's genealogy needs to be considered, and thus only events for the genealogy in question need to be tracked [44–47]. Unfortunately, including arbitrary selection models in coalescent simulations is difficult, because coalescent models rely on the assumption that mutations are independent of the genealogy of the sample - an assumption that is violated in essentially any model including selection. While certain scenarios can be modeled circumventing this problem (such as modeling a selected allele as an additional population connected by migration [48]), the integration of more complex scenarios represents a challenge [48,49].

In the development of both forward and backward simulation tools, a major recent focus has been on modeling positive selection under arbitrary nonequilibrium models. As it is difficult to analytically obtain expectations for patterns of variation under these conditions, simulations become important: these models are indeed tractable to simulate, and both forward [50] and coalescent simulation programs [46] have recently been released that provide better performance under models of arbitrary demographic history. Utilizing today's computational resources, the application of more complex models is feasible, allowing for highly parameterized demographic histories [32,51,52], and permitting direct inference of selection using ABC.

More recently, there is growing awareness of the effects of BGS [53,54] in shaping genomic patterns and on the performance of methods designed for detecting positive selection. Intermediate levels of BGS pose a difficult problem in modeling, at least with coalescent simulators, despite the relative ease of simulating BGS in a forward-simulation framework [40]. While strongly deleterious mutations are purged from the population immediately, and very slightly deleterious mutations (Figure 2) accumulate according to non-neutral dynamics that have yet to be studied in more detail [55].

The growing awareness of the importance of accounting for BGS in future studies is comparable to recent progress in the consideration of demography – where previously it was common to assume an equilibrium population history when performing tests of selection, now demography is commonly considered in such inference. BGS thus appears to be the next frontier to be considered in the development of simulation tools. However, as computing speed and algorithmic developments improve, it should become a priority to develop full inference schemes for the co-estimation of demography and BGS in the future [56]. In particular, direct estimation of the DFE in both experimental and natural populations would lead to new insights into the evolutionary process. Variation of estimated parameters over the genome, and when applicable, over time, can lead to a deeper understanding of how the dynamics of demography, recombination, selection, and mutation drive evolution in natural populations. Complementary to such inference is the potential for validation of statistical methods under known demographic and selection models via experimental evolution studies.

# Experimental evolution as a new source of information

Through advances in sequencing techniques and bioengineering, experimental evolution approaches (see Box 1) have become a flourishing area of development in evolutionary biology, catching increasing interest from the field and enabling a large-scale evaluation of mutational effects that was previously unthinkable. Two general types of approaches dominate this area, one relying on the accumulation of naturally arising mutations over many generations, and one based on the simultaneous introduction and competition of hundreds or thousands of engineered

Table 2. A selection of commonly used simulation tools and included features<sup>a</sup>

Name	ms	msHot	fastsimcoal	msms	SFSCode	SLiM	SimuPop
Refs	[44]	[45]	[47]	[46]	[40]	[50]	[38]
Type of simulator	Coalescent	Coalescent	Coalescent	Coalescent	Forward	Forward	Forward
Recombination	Υ	HotSpots	Υ	Υ	Υ	Υ	Υ
Migration	Υ	Υ	Υ	Υ	Υ	Υ	Υ
Hard sweeps	N	N	N	Υ	Υ	Υ	N
Soft sweeps	N	N	N	Υ	Υ	Υ	N
Linked selection	N	N	N	N	Υ	Υ	Υ
DFE <sup>b</sup>	N	N	N	N	Υ	Υ	Υ
Full chromosomes	N	N	Υ	N	N°	Υ	Υ
Time samples	N	N	Υ	Υ	Υ	Υ	N

<sup>&</sup>lt;sup>a</sup>Y = supported; N = not available.

<sup>&</sup>lt;sup>b</sup>Support of explicit (but not necessarily arbitrary) DFE

<sup>&</sup>lt;sup>c</sup>Not a general restriction, but for performance reasons whole chromosomes are impractical.

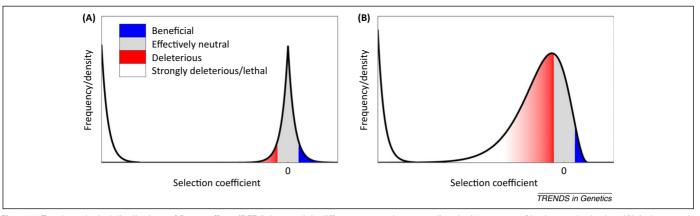


Figure 2. Two hypothetical distributions of fitness effects (DFEs) that result in different expectations regarding the importance of background selection. (A) A sharp mode around neutrality (here represented as exponential decay in both the negative and positive direction), as frequently assumed in population genomic studies of the DFE (e.g., [69]), results in little potential for background selection (BGS; indicated by the red area under the curve). (B) Experimental evolution studies of the DFE suggest a wider mode of the DFE around neutrality that is skewed (and sometimes shifted) towards deleterious mutations (here represented by a shifted negative gamma distribution, as suggested in [70]). This results in a much higher proportion of slightly deleterious mutations that can contribute to BGS. The intensity of the red shading indicates which area under the curve is most likely to contribute to relevant effects of BGS (see also [66]). The second, strongly deleterious mode of the DFE is depicted as exponentially distributed. This is an arbitrary choice, because the accuracy of currently available methods is not sufficient to detect its actual shape.

mutations. Below, we highlight three implications emerging from recent studies that we believe are particularly relevant for those interested in selection inference.

Firstly, approaches using engineered mutations in viruses or microbes allow for a systematic picture of the DFE of all possible point mutations in a chosen region of a gene, hence enabling an assessment of the proportions of beneficial, neutral, deleterious, and even lethal mutations

#### Box 1. Experimental evolution approaches

In traditional experimental evolution studies, a lab strain of an organism (typically a virus, bacterium, microbe, or fly) is exposed to a new and potentially challenging environment, and after a number of generations, its fitness (often measured as relative growth rate in competition with the original wild type) is evaluated. The first laboratory evolution experiment of that kind was performed by William Henry Dallinger between 1880 and 1886 – growing microbes in an incubator under increasing temperature, and observing that they evolved to tolerate previously lethal conditions (reviewed in [71]). Today's approaches fall into two categories that diverge from the original experiment in their increasing use of biotechnology and next generation sequencing:

- (i) Mutation accumulation experiments. With the most prominent of these being Lenski's long term selection experiment in *E. coli* (e.g., [72]) that has been running for more than 50 000 generations, these experiments are very similar to the one explained above, now with the added perspective of whole genome sequencing that enables identification of mutations accumulated or segregating, and with robots enabling maintenance of hundreds of replicates (reviewed in [73,74]).
- (ii) Mutagenesis experiments. Here, hundreds or thousands of individuals are created that carry only one or a few (random or specific) mutations, usually only in a single gene (reviewed in [75]). These are grown for a short period of time and fitness is assessed either by sequencing and estimation of relative growth rate (e.g., [58]), or by assessing other fitness-related phenotypes (e.g., [76,77]).

Advantages and limitations. Experimental evolution studies offer not only new insights into mutational effects, but also a systematic way of testing population genetic models and theories under controlled conditions – hence bridging theory and nature. They are ideally suited to inform us about expected statistical patterns of the DFE and to study resistance evolution. By contrast, they are restricted to certain organisms, and it is generally impossible to reproduce the ecology of a natural environment in the lab.

(e.g., [57,58]). The general picture thus far confirms the one proposed by the nearly-neutral theory [59], postulating a bimodal DFE with one mode centered at wild type-like, and another at strongly deleterious fitness. Notably, however, most studies identify a wide wild type-like mode strongly skewed towards deleterious selection coefficients (Figure 2). This suggests the heavy influence of effective population size in dictating the neutral class of sites, and further highlights the importance of considering the effects of BGS. Integrating our emerging knowledge of the shape of the DFE into simulation tools will be an important step towards better selection inference.

Secondly, as the scale of evolution experiments increases, it is more feasible to study not only the effects of single, but also those of double and multiple-step mutations [78]. A pattern of common epistasis emerges (in particular within genes or pathways), in which the combined effects of mutations are very difficult to predict from the measured singular effects of a given mutation. This might result in the 'missing heritability' commonly observed in genome-wide association studies [60]. In particular, this suggests that the genetic background must be considered when trying to detect single alleles under selection.

Thirdly, although lab environments cannot reproduce natural conditions (e.g., [61]), there are numerous cases in which laboratory evolution experiments have re-identified known antibiotic or antiviral resistance mutations from natural populations, even if grown under quite unnatural conditions (e.g., [32]), or have provided other important results regarding the evolution of resistance (reviewed in [62]) – indicating that experimental evolution results are indeed informative for inference about natural populations. In addition, experimental evolution approaches allow for the study of the effects of the same mutations in different environments. Several such studies have indicated pervasive costs of adaptation, including both mutations of large effect (reviewed in [63,64]) and of small effect [65]. These results suggest that if, upon a change of the environment, adaptation is common from standing genetic variation instead of new mutations, the variation may likely be rare and segregating under mutation-selection balance [66].

#### A roadmap for improved selection inference

Despite > 100 years of population genetic research, major advances in our understanding of genetics, and huge amounts of polymorphism data across organisms and populations, many of the initial challenges that Fisher and Wright faced persist today. However, it is fair to say that many of the necessary next steps are clear, and encouragingly, many of the essential pieces appear ready to be assembled to make such advances. We suggest the following as guidelines for moving the field forward:

- (i) Continued improvements of simulation tools are allowing increasingly relevant models to be considered – but computationally efficient programs capable of simulating a wide range of complex models represent an ongoing challenge. Relatedly, simulations should become an obligatory instrument to assess the power of inference, to test new methods, and to consider alternative models.
- (ii) ABC is a fast and efficient inference method that is likely the way forward for co-estimating demography and BGS with positive selection. However, the identification of summary statistics that are capable of distinguishing between these processes, particularly given that there may not be genomic sites impacted by only a single one of these effects, remains a pressing challenge.
- (iii) Results from experimental evolution complement population genomics by yielding important insights into the DFE and the extent of epistatic interactions. An explicit and realistic shape of the DFE can now be incorporated both into simulation algorithms and in inference methods applied to genomic polymorphism data in natural populations.

This opinion piece, therefore, represents a roadmap for improved selection inference – utilizing novel theory/method development combined with the tremendous amount of data currently being made available via next-generation sequencing. As such, we may for example draw information from experimental evolution studies, and leverage it against time-sampled population data allowing for entire mutational trajectories to be studied within the context of the underlying distribution of fitness effects. With these theoretical and experimental tools, population genetics is indeed on the verge of important breakthroughs in not only our general understanding of the process of adaptation, but also in the ability of evolutionary analysis to provide important insights to related fields ranging from ecology to medicine.

# **Acknowledgements**

Financial support for this work was provided by grants from the Swiss National Science Foundation and the European Research Council to J.D.J. The authors are grateful to H. Shim for help with the comparison of multitime point methods, and to K. Irwin for a careful reading of the manuscript.

#### Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tig.2014.09.010.

#### References

- 1 Nielsen, R. (2005) Molecular signatures of natural selection. Annu. Rev. Genet. 39, 197–218
- 2 Jensen, J.D. et al. (2007) Approaches for identifying targets of positive selection. Trends Genet. 23, 568–577
- 3 Kumar, S. et al. (2012) Statistics and truth in phylogenomics. Mol. Biol. Evol. 29, 457–472
- 4 Lawrie, D.S. and Petrov, D.A. (2014) Comparative population genomics: power and principles for the inference of functionality. *Trends Genet.* 30, 133–139
- 5 Fisher, R.A. and Ford, E.B. (1947) The spread of a gene in natural conditions in a colony of the moth *Panaxia dominula L. Heredity* 1, 143-174
- 6 Clegg, M.T. et al. (1976) Dynamics of correlated genetic systems. I. Selection in the region of the Glued locus of Drosophila melanogaster. Genetics 83, 793–810
- 7 Lewontin, R.C. and Krakauer, J. (1973) Distribution of gene frequency as a test of the theory of the selective neutrality of polymorphisms. Genetics 74, 175–195
- 8 Beaumont, M.A. and Balding, D.J. (2004) Identifying adaptive genetic divergence among populations from genome scans. *Mol. Ecol.* 13, 969–980
- 9 Foll, M. and Gaggiotti, O. (2008) A genome-scan method to identify selected loci appropriate for both dominant and codominant markers: a Bayesian perspective. *Genetics* 180, 977–993
- 10 Vitalis, R. et al. (2014) Detecting and measuring selection from gene frequency data. Genetics 196, 799–817
- 11 Kim, Y. and Stephan, W. (2002) Detecting a local signature of genetic hitchhiking along a recombining chromosome. Genetics 160, 765–777
- 12 Jensen, J.D. *et al.* (2005) Distinguishing between selective sweeps and demography using DNA polymorphism data. *Genetics* 170, 1401–1410
- 13 Nielsen, R. et al. (2005) Genomic scans for selective sweeps using SNP data. Genome Res. 15, 1566–1575
- 14 Kim, Y. and Nielsen, R. (2004) Linkage disequilibrium as a signature of selective sweeps. Genetics 167, 1513–1524
- 15 Voight, B.F. et al. (2006) A map of recent positive selection in the human genome. PLoS Biol. 4, e72
- 16 Pavlidis, P. et al. (2013) SweeD: likelihood-based detection of selective sweeps in thousands of genomes. Mol. Biol. Evol. 30, 2224–2234
- 17 Ferrer-Admetlla, A. et al. (2014) On detecting incomplete soft or hard selective sweeps using haplotype structure. Mol. Biol. Evol. 31, 1275–1291
- 18 Thornton, K.R.  $et\ al.$  (2007) Progress and prospects in mapping recent selection in the genome.  $Heredity\ 98,\ 340-348$
- 19 Crisci, J.L. et al. (2012) Recent progress in polymorphism-based population genetic inference. J. Hered. 103, 287–296
- 20 Gutenkunst, R.N. et al. (2009) Inferring the joint demographic history of multiple populations from multidimensional SNP frequency data. PLoS Genet. 5, e1000695
- 21 Excoffier, L. et al. (2013) Robust demographic inference from genomic and SNP data. PLoS Genet. 9, e1003905
- 22 Charlesworth, B. (2013) Background selection 20 years on: the Wilhelmine E. Key 2012 invitational lecture. J. Hered. 104, 161–171
- 23 Li, J. et al. (2012) Joint analysis of demography and selection in population genetics: where do we stand and where could we go? Mol. Ecol. 21, 28–44
- 24 Sunnåker, M. et al. (2013) Approximate Bayesian computation. PLoS Comput. Biol. 9, e1002803
- 25 Bazin, E. et al. (2010) Likelihood-free inference of population structure and local adaptation in a Bayesian hierarchical model. Genetics 185, 587–602
- 26 Barthelmé, S. and Chopin, N. (2014) Expectation propagation for likelihood-free inference. J. Am. Stat. Assoc. 109, 315–333
- 27 Waples, R.S. (1989) A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics* 121, 379–391
- 28 Williamson, E.G. and Slatkin, M. (1999) Using maximum likelihood to estimate population size from temporal changes in allele frequencies. Genetics 152, 755–761
- 29 Anderson, E.C. et al. (2000) Monte Carlo evaluation of the likelihood for Ne from temporally spaced samples. Genetics 156, 2109–2118
- 30 Drummond, A.J. and Rambaut, A. (2007) BEAST: Bayesian evolutionary analysis by sampling trees. BMC Evol. Biol. 7, 214

- 31 Malaspinas, A.-S. et al. (2012) Estimating allele age and selection coefficient from time-serial data. Genetics 192, 599–607
- 32 Foll, M. et al. (2014) Influenza virus drug resistance: a time-sampled population genetics perspective. PLoS Genet. 10, e1004185
- 33 Foll, M. et al. (2014) WFABC: a Wright–Fisher ABC-based approach for inferring effective population sizes and selection coefficients from time-sampled data. Mol. Ecol. Resour. Published online June 11, 2014. (http://dx.doi.org/10.1111/1755-0998.12280)
- 34 Mathieson, I. and McVean, G. (2013) Estimating selection coefficients in spatially structured populations from time series data of allele frequencies. *Genetics* 193, 973–984
- 35 Illingworth, C.J.R. et al. (2012) Quantifying selection acting on a complex trait using allele frequency time series data. Mol. Biol. Evol. 29, 1187–1197
- 36 Tajima, F. (1989) Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics 123, 585–595
- 37 Grossman, S.R. et al. (2010) A composite of multiple signals distinguishes causal variants in regions of positive selection. Science 327, 883–886
- 38 Peng, B. and Kimmel, M. (2005) simuPOP: a forward-time population genetics simulation environment. *Bioinformatics* 21, 3686–3687
- 39 Peng, B. et al. (2007) Forward-time simulations of human populations with complex diseases. PLoS Genet. 3, e47
- 40 Hernandez, R.D. (2008) A flexible forward simulator for populations subject to selection and demography. Bioinformatics 24, 2786–2787
- 41 Padhukasahasram, B. et al. (2008) Exploring population genetic models with recombination using efficient forward-time simulations. Genetics 178, 2417–2427
- 42 Messer, P.W. and Petrov, D.A. (2013) Population genomics of rapid adaptation by soft selective sweeps. *Trends Ecol. Evol.* 28, 659-669
- 43 Kessner, D. and Novembre, J. (2014) forqs: Forward-in-time simulation of recombination, quantitative traits and selection. *Bioinformatics* 30, 576–577
- 44 Hudson, R.R. (2002) Generating samples under a Wright-Fisher neutral model of genetic variation. *Bioinformatics* 18, 337–338
- 45 Hellenthal, G. and Stephens, M. (2007) msHOT: modifying Hudson's ms simulator to incorporate crossover and gene conversion hotspots. *Bioinformatics* 23, 520–521
- 46 Ewing, G. and Hermisson, J. (2010) MSMS: a coalescent simulation program including recombination, demographic structure and selection at a single locus. *Bioinformatics* 26, 2064–2065
- 47 Excoffier, L. and Foll, M. (2011) fastsimcoal: a continuous-time coalescent simulator of genomic diversity under arbitrarily complex evolutionary scenarios. *Bioinformatics* 27, 1332–1334
- 48 Hudson, R.R. (1990) Gene genealogies and the coalescent process. In Oxford Surveys in Evolutionary Biology (Vol. 7) (Futuyma, D. and Antonovics, J., eds), pp. 1–44
- 49 Wakeley, J. (2009) Coalescent Theory, Roberts Publishers
- 50 Messer, P.W. (2013) SLiM: simulating evolution with selection and linkage. Genetics 194, 1037–1039
- 51 Wegmann, D. et al. (2009) Efficient approximate Bayesian computation coupled with Markov chain Monte Carlo without likelihood. Genetics 182, 1207–1218
- 52 Johnson, P.L.F. and Hellmann, I. (2011) Mutation rate distribution inferred from coincident SNPs and coincident substitutions. Genome Biol. Evol. 3, 842–850
- 53 Charlesworth, B. et al. (1993) The effect of deleterious mutations on neutral molecular variation. Genetics 134, 1289–1303
- 54 Comeron, J.M. (2014) Background selection as baseline for nucleotide variation across the *Drosophila* genome. PLoS Genet. 10, e1004434

- 55 Charlesworth, B. et al. (1997) The effects of local selection, balanced polymorphism and background selection on equilibrium patterns of genetic diversity in subdivided populations. Genet. Res. 70, 155–174
- 56 Zeng, K. and Charlesworth, B. (2011) The joint effects of background selection and genetic recombination on local gene genealogies. *Genetics* 189, 251–266
- 57 Sanjuán, R. (2010) Mutational fitness effects in RNA and single-stranded DNA viruses: common patterns revealed by site-directed mutagenesis studies. *Philos Trans. R. Soc. Lond. B: Biol. Sci.* 365, 1975–1982
- 58 Bank, C. et al. (2014) A Bayesian MCMC approach to assess the complete distribution of fitness effects of new mutations: uncovering the potential for adaptive walks in challenging environments. Genetics 196, 841–852
- 59 Ohta, T. (1973) Slightly deleterious mutant substitutions in evolution. Nature 246, 96–98
- 60 Mackay, T.F.C. (2014) Epistasis and quantitative traits: using model organisms to study gene–gene interactions. Nat. Rev. Genet. 15, 22–33
- 61 Kvitek, D.J. and Sherlock, G. (2013) Whole genome, whole population sequencing reveals that loss of signaling networks is the major adaptive strategy in a constant environment. PLoS Genet. 9, e1003972
- 62 Jansen, G. et al. (2013) Experimental evolution as an efficient tool to dissect adaptive paths to antibiotic resistance. Drug Resist. Updat. 16, 96–107
- 63 Andersson, D.I. and Hughes, D. (2010) Antibiotic resistance and its cost: is it possible to reverse resistance? Nat. Rev. Microbiol. 8, 260–271
- 64 Goldhill, D.H. and Turner, P.E. (2014) The evolution of life history trade-offs in viruses. *Curr. Opin. Virol.* 8, 79–84
- 65 Hietpas, R.T. et al. (2013) Shifting fitness landscapes in response to altered environments. Evolution 67, 3512–3522
- 66 Jensen, J.D. (2014) On the unfounded enthusiasm for soft selective sweeps. Nat. Commun. 5, 5281
- 67 Ewens, W.J. (2004) Mathematical Population Genetics 1: Theoretical Introduction, Springer, New York
- 68 Bollback, J.P. et al. (2008) Estimation of 2Nes from temporal allele frequency data. Genetics 179, 497–502
- 69 Eyre-Walker, A. and Keightley, P.D. (2009) Estimating the rate of adaptive molecular evolution in the presence of slightly deleterious mutations and population size change. Mol. Biol. Evol. 26, 2097–2108
- 70 Martin, G. and Lenormand, T. (2006) A general multivariate extension of Fisher's geometrical model and the distribution of mutation fitness effects across species. *Evolution* 60, 893–907
- 71 Davenport, C.B. and Castle, W.E. (1895) Studies in morphogenesis, III.
  On the acclimatization of organisms to high temperatures. Archiv für Entwicklungsmechanik der Organismen 2, 227–249
- 72 Lenski, R.E. et al. (1991) Long-term experimental evolution in Escherichia coli. I. Adaptation and divergence during 2000 generations. Am. Nat. 138, 1315–1341
- 73 Barrick, J.E. and Lenski, R.E. (2013) Genome dynamics during experimental evolution. *Nat. Rev. Genet.* 14, 827–839
- 74 Desai, M.M. (2013) Statistical questions in experimental evolution. J. Stat. Mech. 2013, P01003
- 75 Fowler, D.M. and Fields, S. (2014) Deep mutational scanning: a new style of protein science. Nat. Methods 11, 801–807
- 76 Jacquier, H. et al. (2013) Capturing the mutational landscape of the beta-lactamase TEM-1. Proc. Natl. Acad. Sci. U.S.A. 110, 13067–13072
- 77 Melamed, D. et al. (2013) Deep mutational scanning of an RRM domain of the Saccharomyces cerevisiae poly(A)-binding protein. RNA 19, 1537–1551
- 78 Bank, C., et al. Molecular Biology and Evolution. http://dx.doi.org/ 10.1093/molbev/msu30, in press.